



# Screening Of Beneficial Microorganisms To Control Root Rot Of Olive Trees

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## **INTRODUCTION**

## **EXPERIMENTAL SECTION**

- *Part 1: Screening of antagonistic bacteria for the biocontrol of olive root rot*
- *Part 2: Use of Trichoderma spp., for the management of olive root rot*

## **CONCLUSION & PERSPECTIVES**

# Introduction

## IMPORTANCE OF THE OLEICULTURE SECTOR

**23.6 M ha**

World olive-growing surface (**2022**)



**1.2 M ha**

Olive-growing area in Morocco

**1.9 M Tons**

Average production of olives



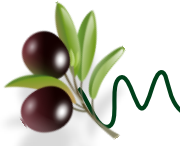
**50 M**

working days per year

Contribution agricultural economy & employment



# Introduction



## PHYTOSANITARY PROBLEMS OF OLIVE TREES

**Verticillium wilt**

*Verticillium dahliae*



**Cycloconium leaf spot**

*Spilocaea oleaginea*



**Anthracnose**  
*Colletotrichum spp.,*



**Tuberculosis**

*Pseudomonas savastanoi*



# Introduction



## ROOT ROT DISEASE OF OLIVE TREES

Oomycetes, *Pythium* genera, have been described as agents that induce:



**Disease conditions :** heavy soils, excessive soil moisture, and poor drainage



Affect **olive nurseries & commercial groves**

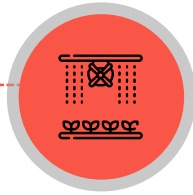


# Introduction



## CONTROL STRATEGIES

1



### Cultural Practices

Prophylaxis/sanitation practices are preventive measures that eliminate stress factors that predispose trees to attack by pathogens.

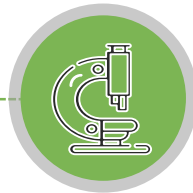
2



### Chemical Control

Use of active fungicidal molecules to control plant pathogens.

3



### Biological Control

Use of biological agents to control plant pathogens and establish an eco-friendly ecosystem.

# Introduction

## OBJECTIVES

**Screening of beneficial microorganisms for the management of root rot in olive trees caused by *Pythium schmitthenneri***





*Part 1: Screening of  
antagonistic bacteria for the  
biocontrol of olive root rot*



# Methodology



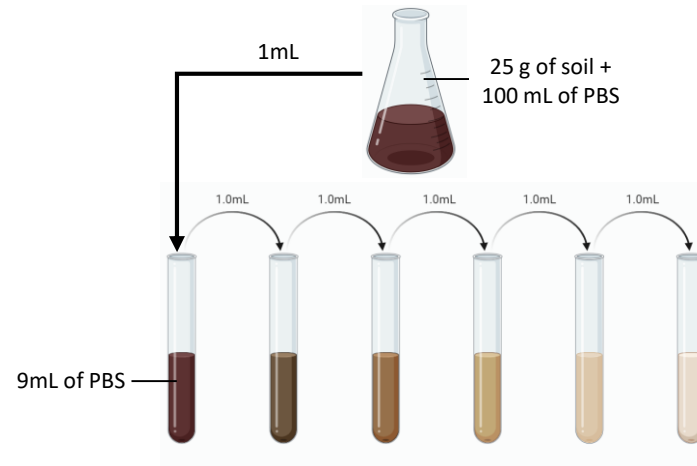
Samples

Olive Rhizosphere:

- ❖ Fes
- ❖ Taza
- ❖ Taourirt

Isolation

Olive Rhizosphere



Incubation & purification

Conservation

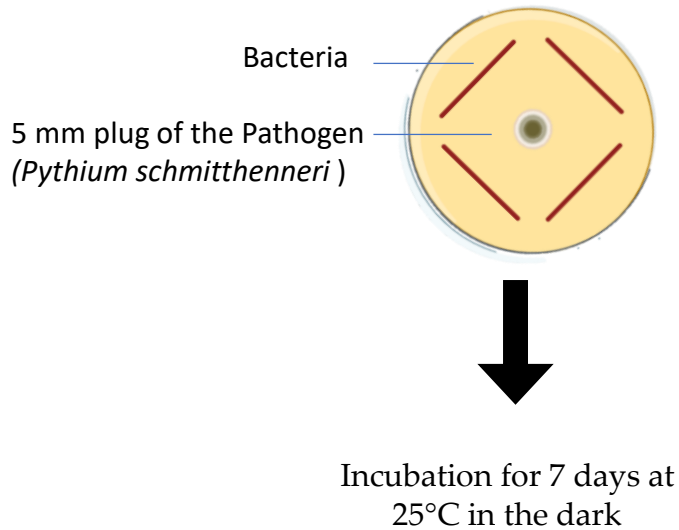
Cryoconservation  
(-80°C)

# Methodology

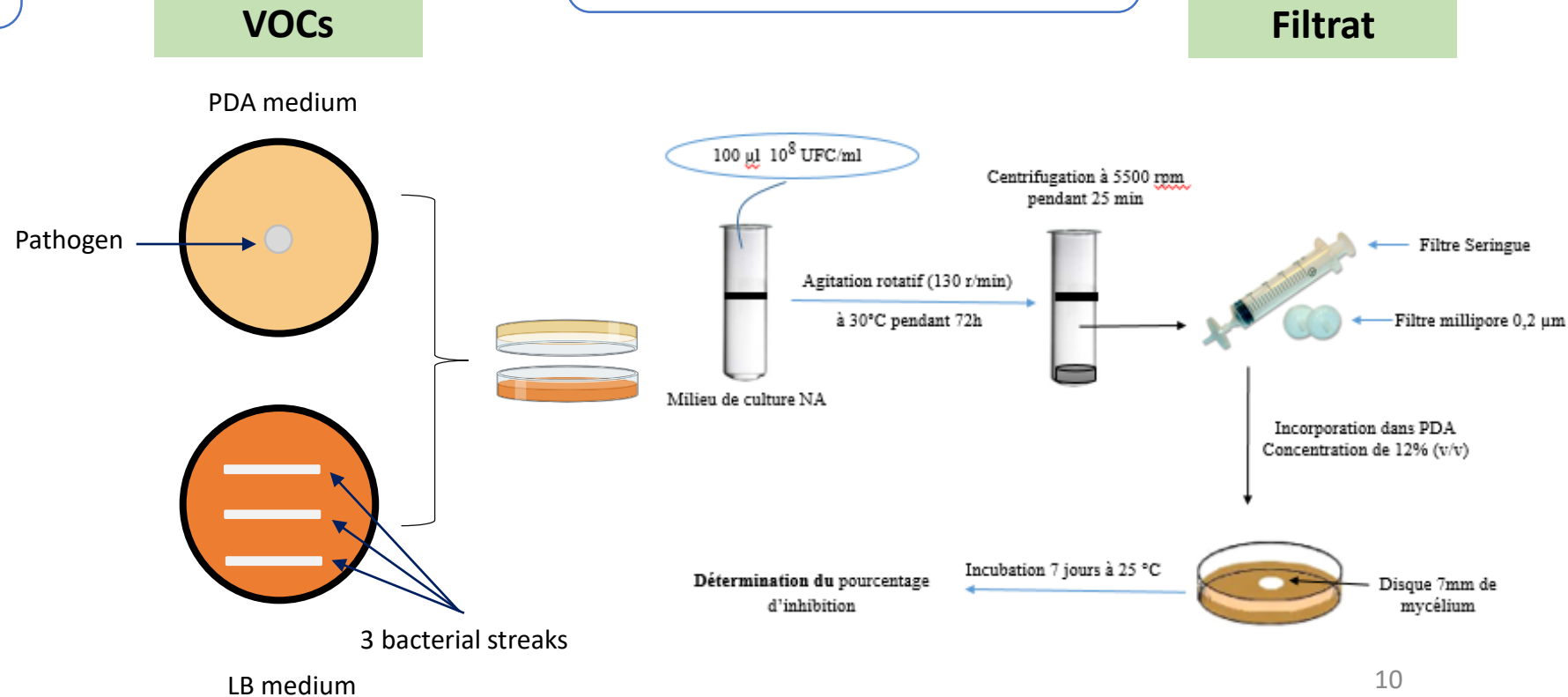
## Valorisation

### Search for bacterial antagonism

#### Screening & Dual culture



#### Indirect confrontations



# Methodology

## Valorisation

### Bacterial DNA extraction

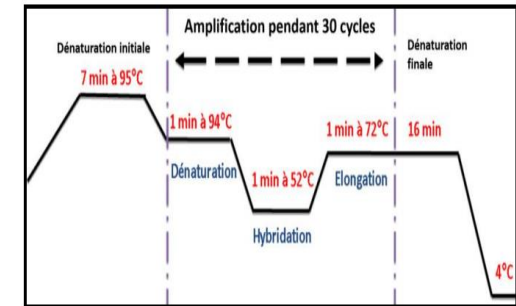
#### Bioneer MagListo TM 5M DNA extraction kit



### Molecular identification

- DNA amplification  
16s rDNA gene

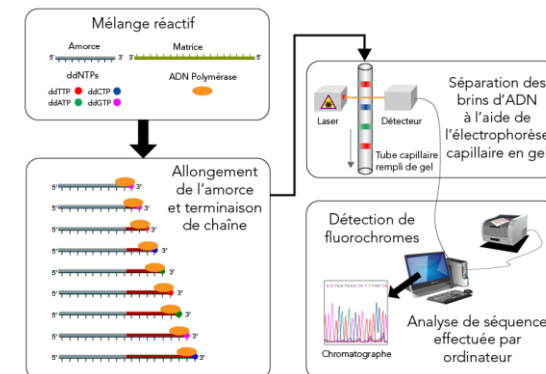
| Primer | Sequence                    |
|--------|-----------------------------|
| FD1    | 5'-AGAGTTTGATCCTGGCTCAG-3'  |
| RP2    | 5'-ACGGCTACCTTGTTACGACTT-3' |



- Amplicon revelation

Evolution of DNA amplification by agarose gel electrophoresis

- Sequencing  
Sanger technology



# Methodology

## Biochemical characterization

### Production of lytic enzymes

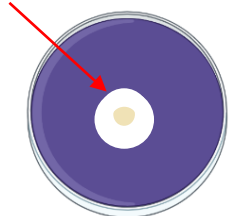
#### Amylase

Starch-based medium



Lugol (0,2% KI)  
12, 2%

clear halos surrounding the colonies



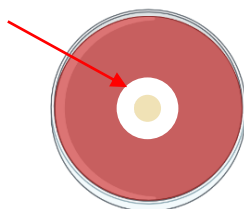
#### Cellulase

CMC-based medium



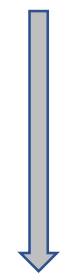
Congo red (0,1%)

clear halos surrounding the colonies

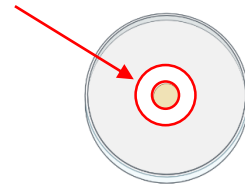


#### Protease

skim milk-based medium



clear halos surrounding the colonies

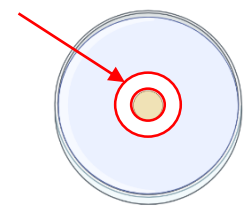


#### Chitinase

Chitine-based medium



clear halos surrounding the colonies



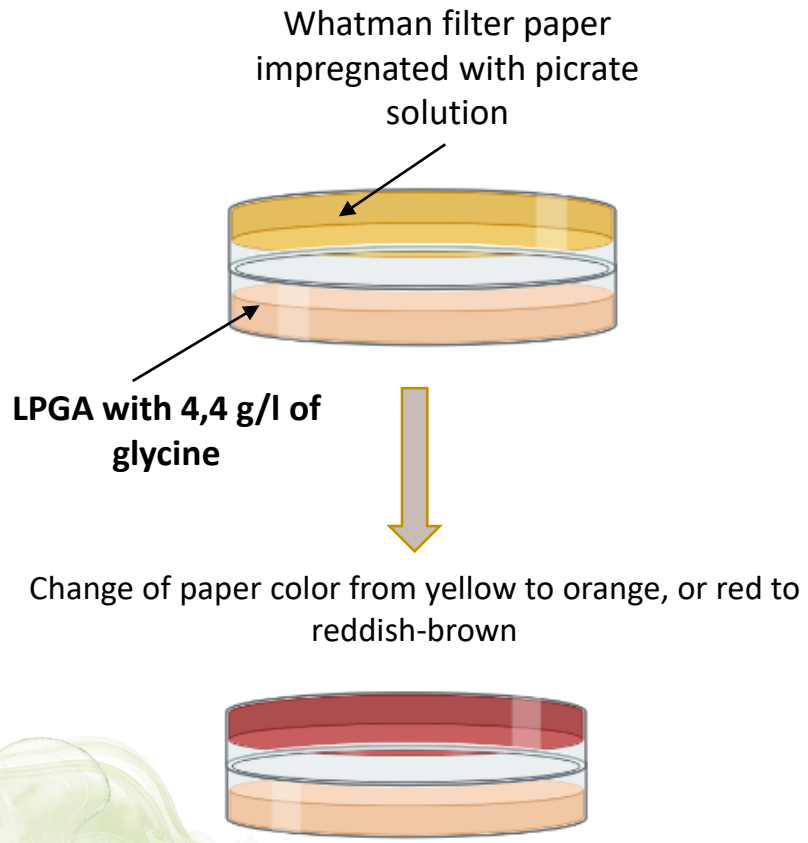
# Methodology



## Biochemical characterization

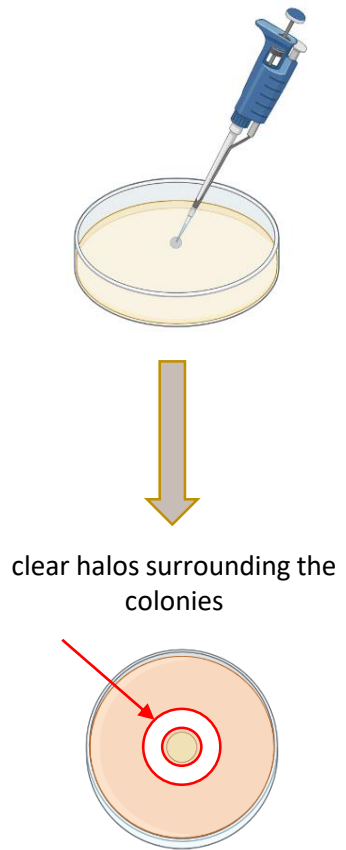
### • Volatile molecules production

#### HCN production

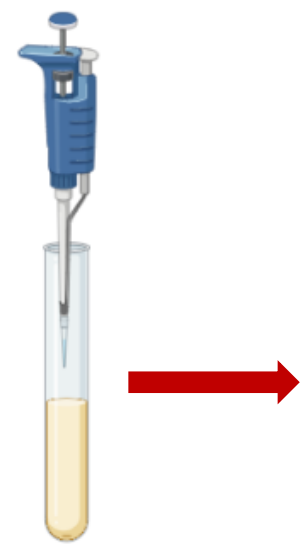


### • Promotion of plant growth

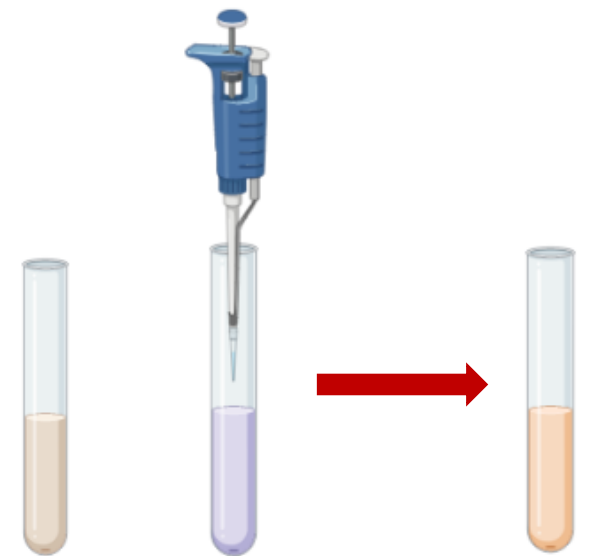
#### Phosphate solubilization



#### IAA production



#### Siderophore secretion



# Methodology

## Application

### In vivo assay

#### 1 Soil sterilization

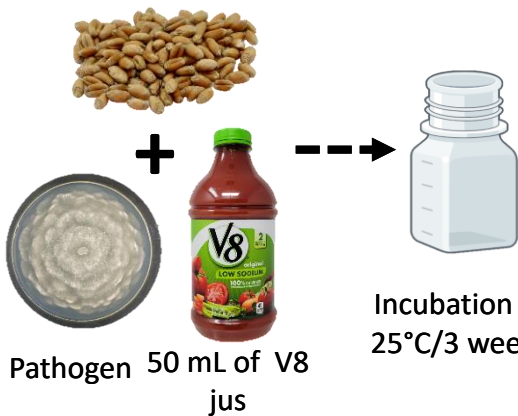


- 121 °C
- 90 min (2 times)



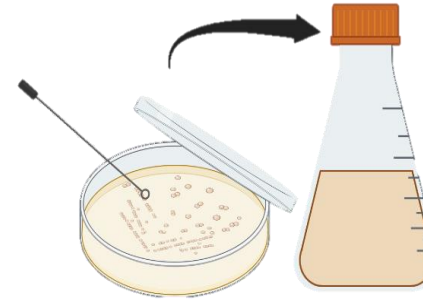
#### 3 Inoculum preparation

50 g of wheat seeds



#### 2 Treatment preparation

200 mL Bacterial suspension



#### 4 In vivo assay

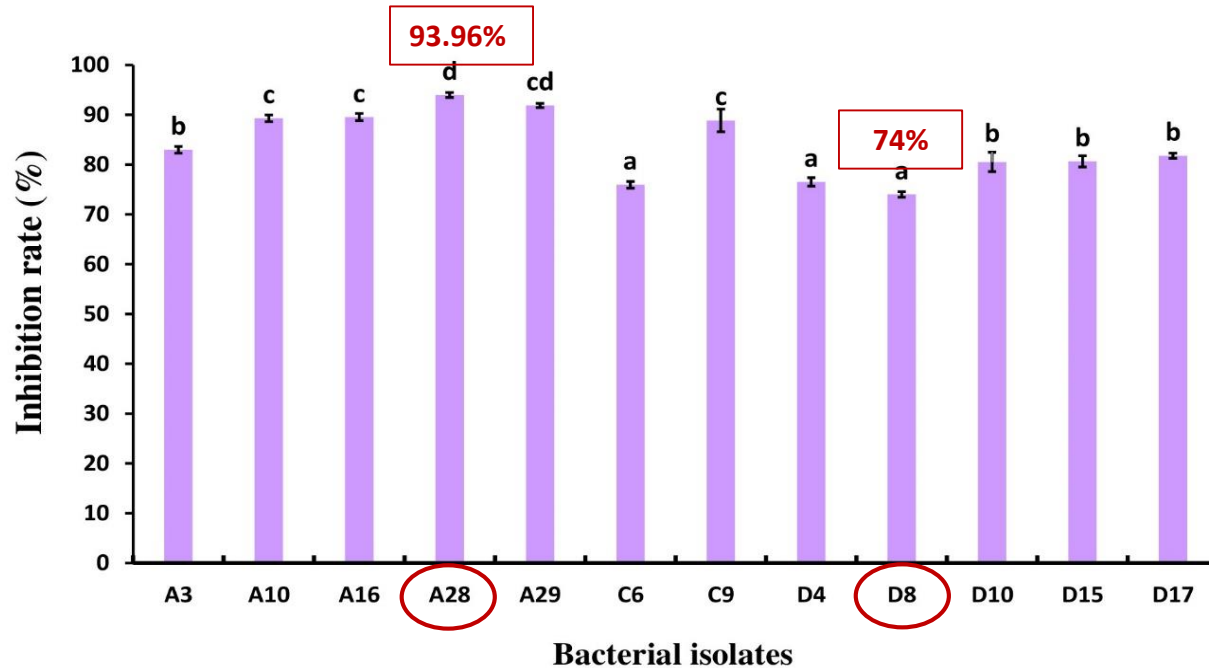


3 months  
Under greenhouse  
condition

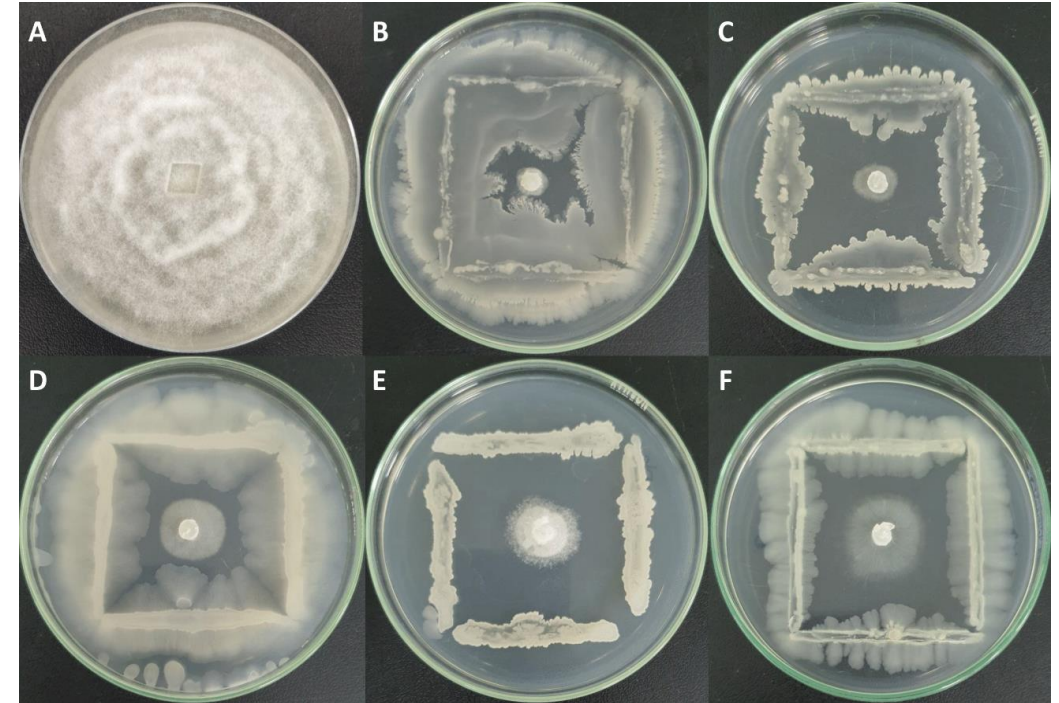
# Results



## Direct antagonistic activity



Inhibition rate (%) of *Pythium schmitthenneri* mycelial growth by twelve selected bacterial isolates from the olive rhizosphere.



Double culture trial showing the antagonistic potential of selected bacteria against *P. schmitthenneri* on PDA medium following six days of incubation at 25°C. (A): untreated control; (B): A28; (C): C9; (D): A3; (E): D17; (F): C6.

# Results

## Indirect confrontations

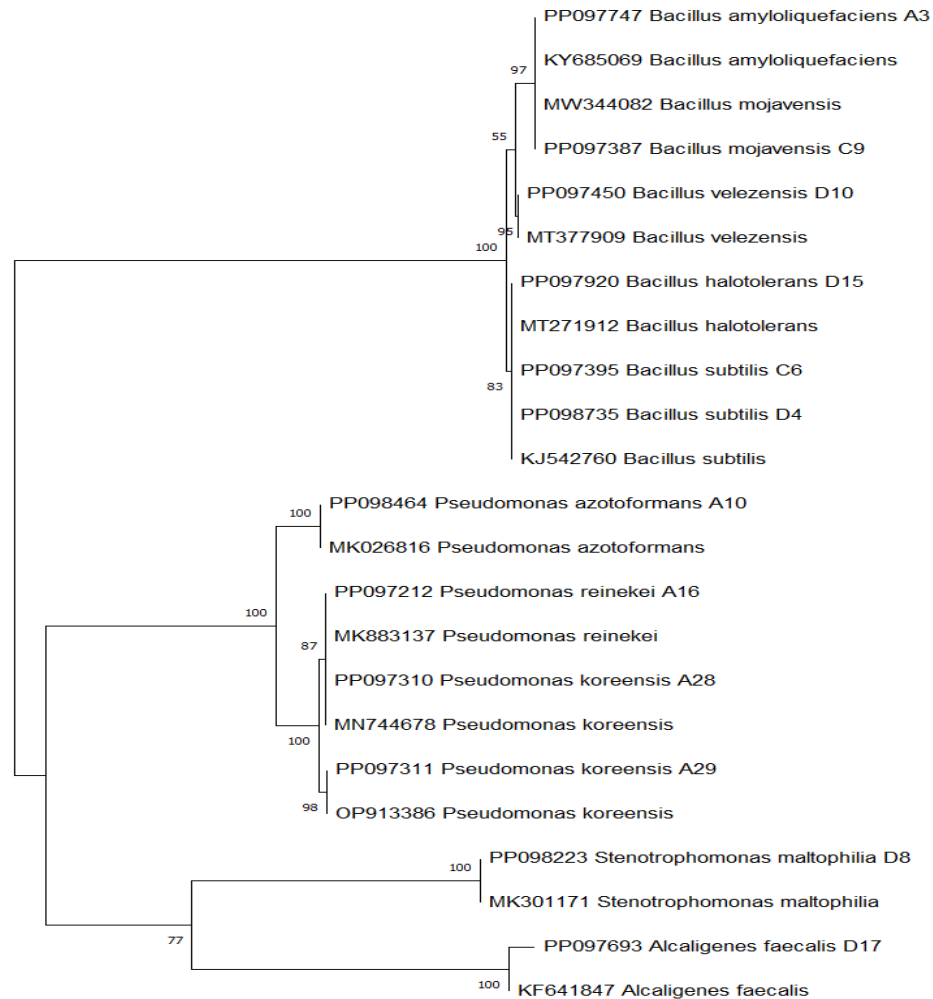
Effects of bacterial volatile organic compounds (VOCs) and cell-free culture filtrates on the growth of *P. schmitteneri* mycelia following 6 days incubation at 25°C.

| Isolate code | Origine  | Species                     | Accession number | VOCs                        | Cell-free filtrate         |
|--------------|----------|-----------------------------|------------------|-----------------------------|----------------------------|
| A3           | Taza     | <i>B. amyloliquefaciens</i> | PP097747         | 31.30 ± 0.43 <sup>a</sup>   | 75.48 ± .55 <sup>d,e</sup> |
| A10          | Taza     | <i>P. azotoformans</i>      | PP098464         | 41.06 ± 1.04 <sup>b</sup>   | 75.78 ± .68 <sup>d,e</sup> |
| A16          | Taza     | <i>P. reinekei</i>          | PP097212         | 68.28 ± 1.31 <sup>f</sup>   | 59.38 ± .72 <sup>a</sup>   |
| A28          | Taza     | <i>P. koreensis</i>         | PP097310         | 89.65 ± 1.06 <sup>i</sup>   | 85.55 ± .51 <sup>g</sup>   |
| A29          | Taza     | <i>P. koreensis</i>         | PP097311         | 80.95 ± 1.38 <sup>j</sup>   | 87.78 ± .88 <sup>h</sup>   |
| C6           | Taourirt | <i>B. subtilis</i>          | PP097395         | 75.20 ± 0.90 <sup>g</sup>   | 74.04 ± .41 <sup>d</sup>   |
| C9           | Taourirt | <i>B. mojavensis</i>        | PP097387         | 55.77 ± 1.64 <sup>e</sup>   | 76.26 ± .64 <sup>e</sup>   |
| D4           | Fez      | <i>B. subtilis</i>          | PP098735         | 45.61 ± 1.18 <sup>c</sup>   | 70.77 ± .63 <sup>c</sup>   |
| D8           | Fez      | <i>S. maltophilia</i>       | PP098223         | 44.31 ± 0.90 <sup>b,c</sup> | 63.62 ± .51 <sup>b</sup>   |
| D10          | Fez      | <i>B. velezensis</i>        | PP097450         | 55.50 ± 1.67 <sup>e</sup>   | 82.29 ± .66 <sup>f</sup>   |
| D15          | Fez      | <i>B. halotolerans</i>      | PP097920         | 42.72 ± 1.32 <sup>b,c</sup> | 58.79 ± .90 <sup>a</sup>   |
| D17          | Fez      | <i>A. faecalis</i>          | PP097693         | 49.25 ± .7 <sup>d</sup>     | 70.17 ± .59 <sup>c</sup>   |



# Results

## Molecular identification



- *Bacillus subtilis* C6 & D4
- *Bacillus amyloliquefaciens* A3
- *Bacillus mojavensis* C9
- *Bacillus velezensis* D10
- *Bacillus halotolerans* D15
  
- *Pseudomonas koreensis* A28 & A29
- *Pseudomonas reinekei* A16
- *Pseudomonas azotoformans* A10
  
- *Alcaligenes faecalis* D17
  
- *Stenotrophomonas maltophilia* D8

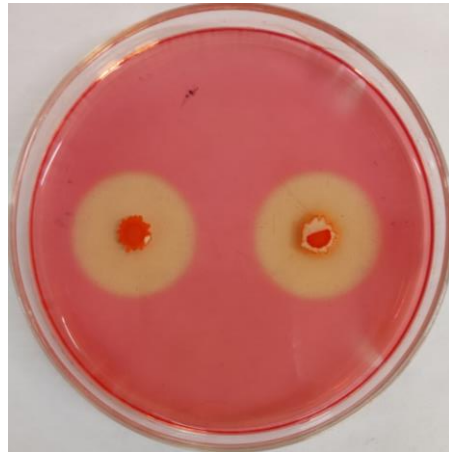


# Results

## Biochemical characterization

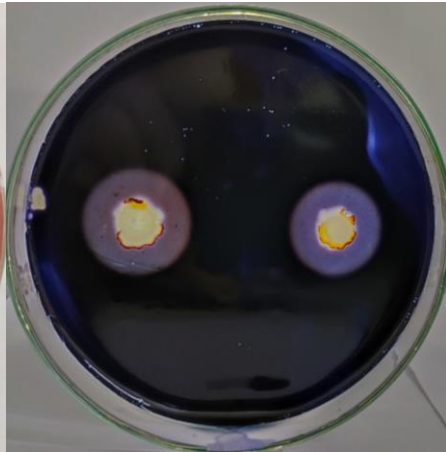
### Production of lytic enzymes

Cellulase



8 isolates

Amylase



8 isolates

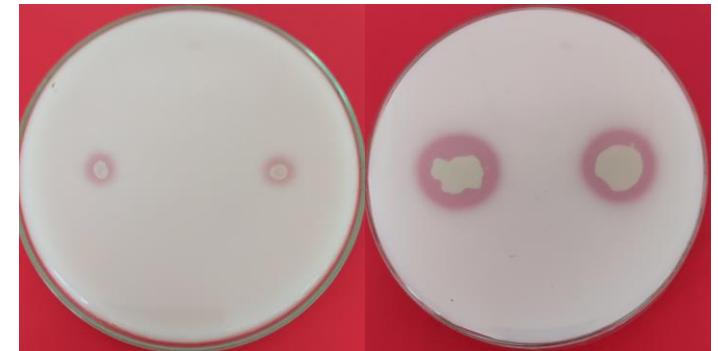
Protease



12 isolates

### Promotion of plant growth

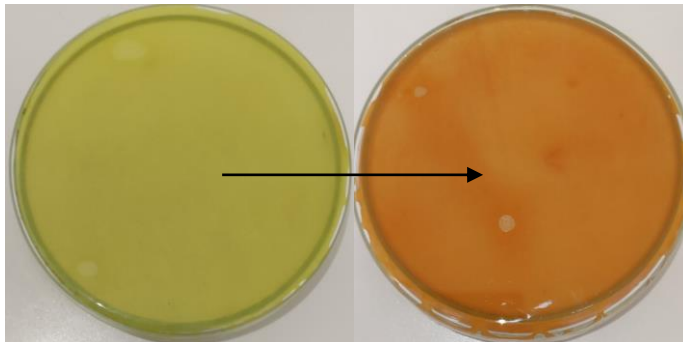
Phosphate solubilization



5 isolates

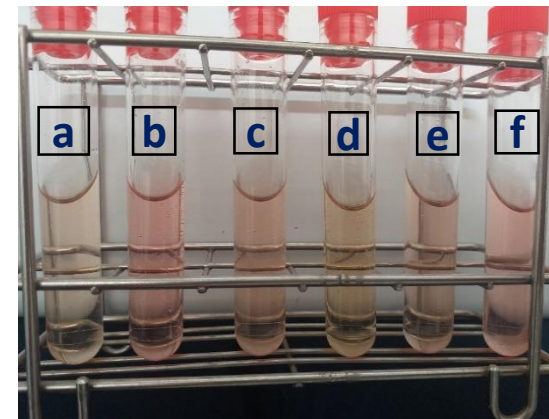
### Volatile molecules production

HCN Production



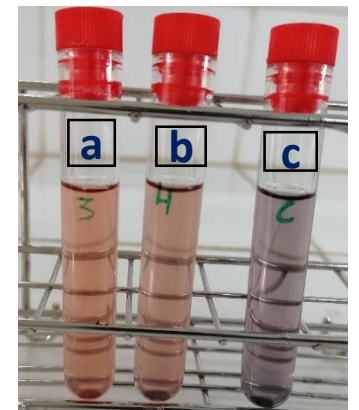
8 isolates

IAA Production



10 isolates

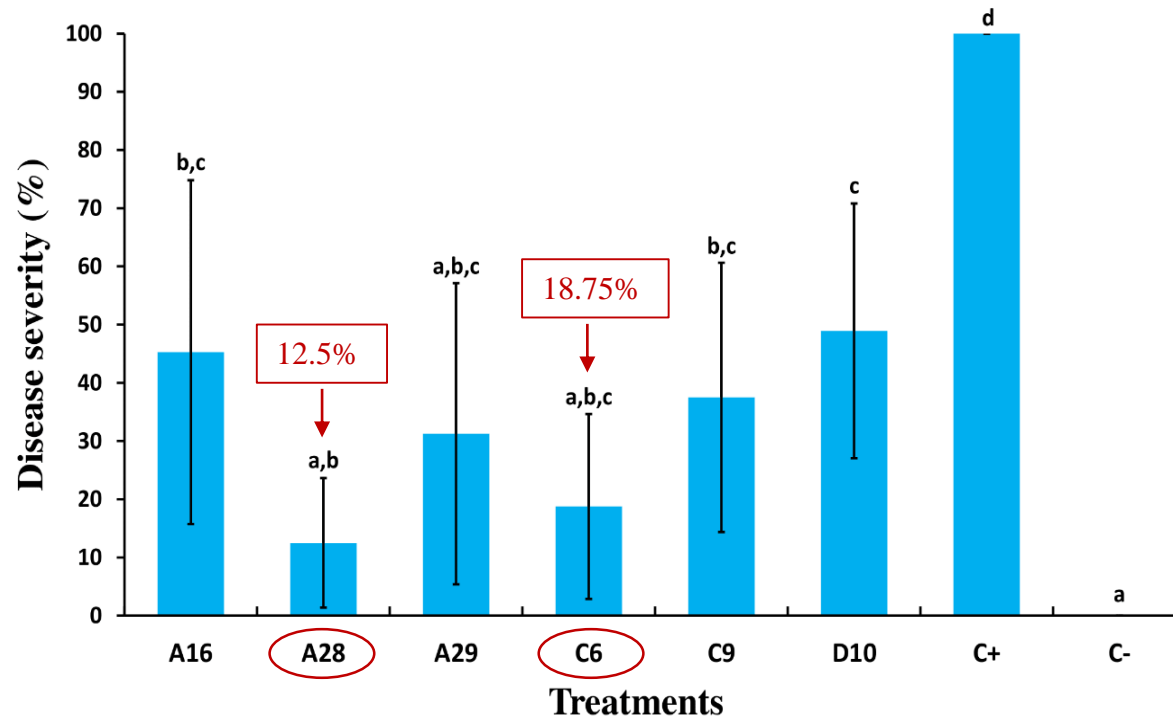
Siderophore secretion



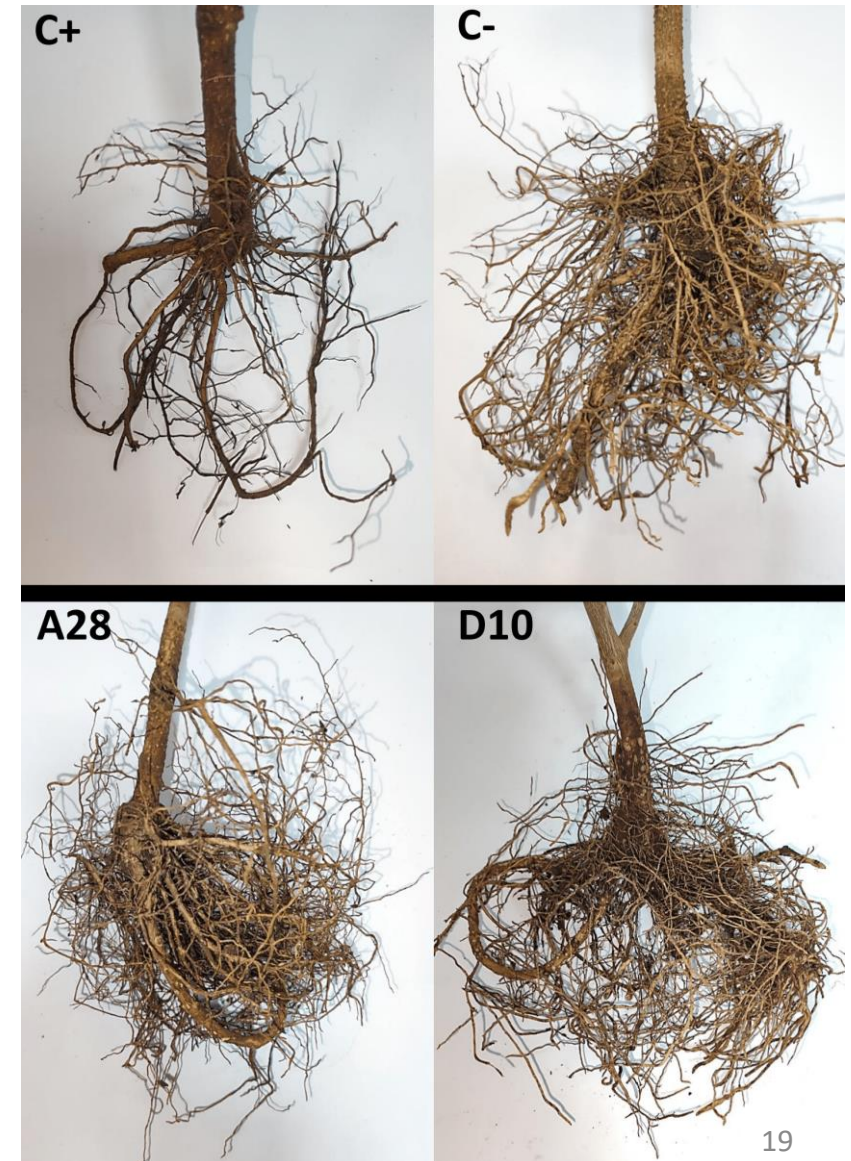
9 isolates

# Results

## Application



- ❖ *Pseudomonas Koreensis* A28 and *Bacillus subtilis* C6 exhibited higher effectiveness in disease suppression in comparison to the positive control

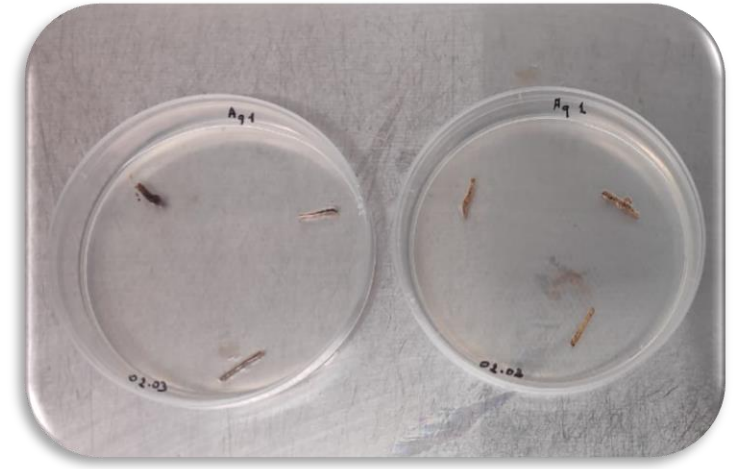
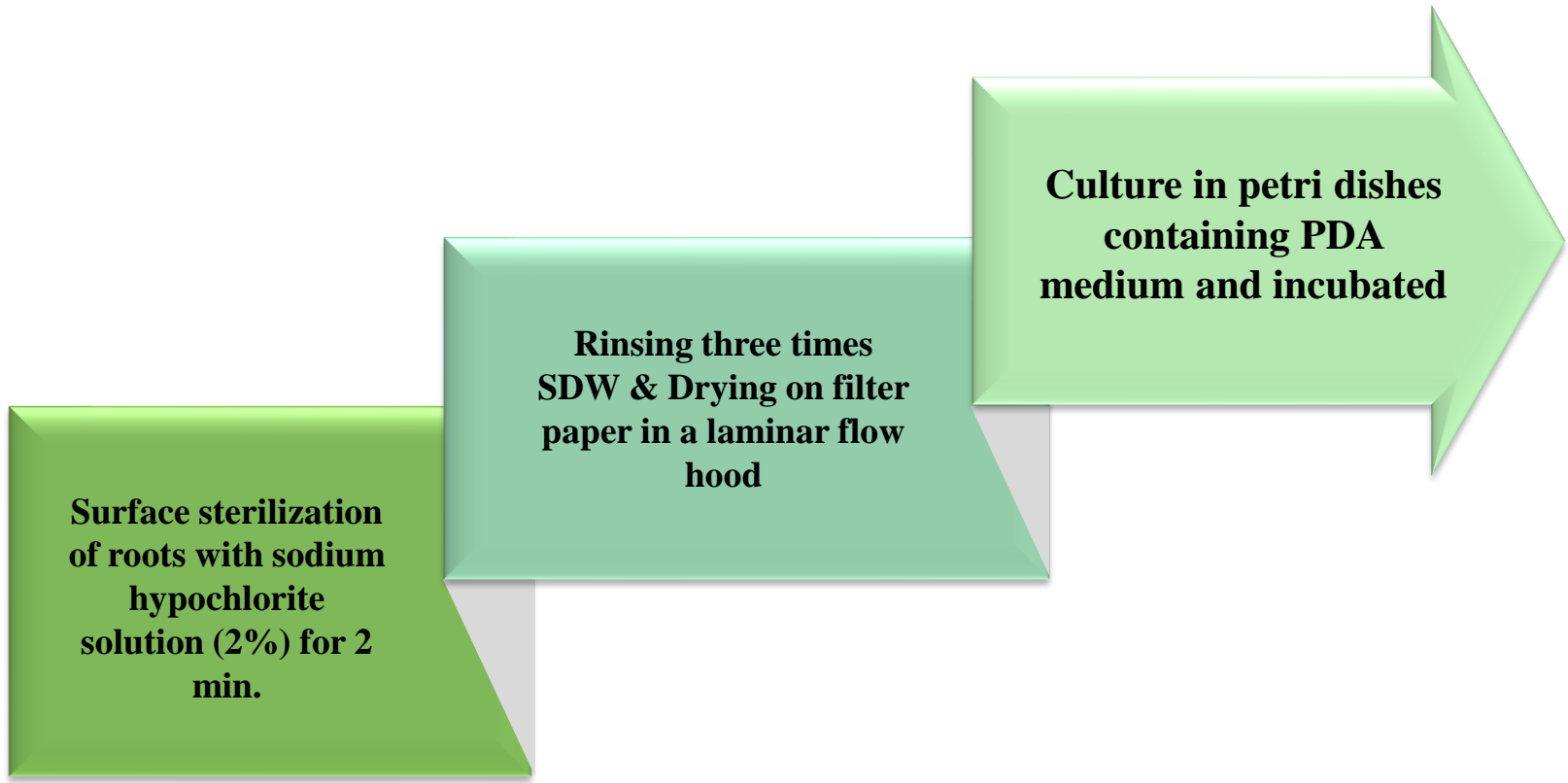




*Part 2: Use of Trichoderma  
spp., for the management of  
olive root rot*

# Methodology

## Isolation



Incubation  
25°C/5 days

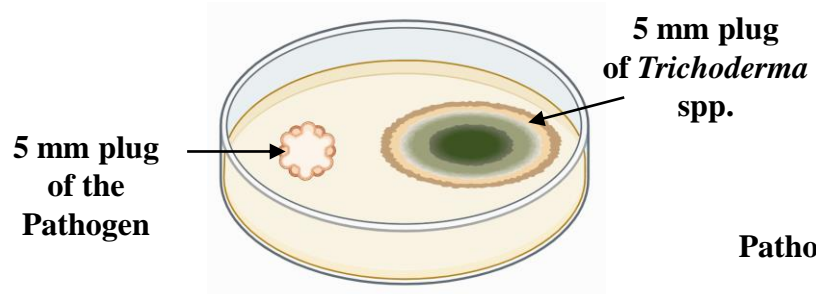
Purification

# Methodology

## Valorisation

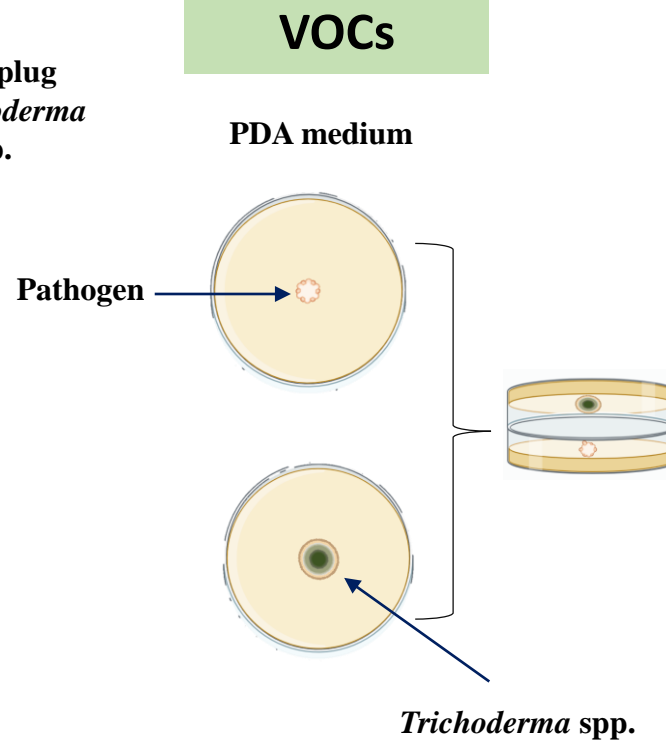
Search for effective *Trichoderma* isolates

### Screening & Dual culture



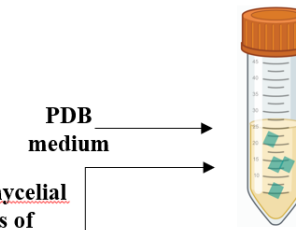
Incubation for 6 days at 25°C in the dark

### Indirect confrontations



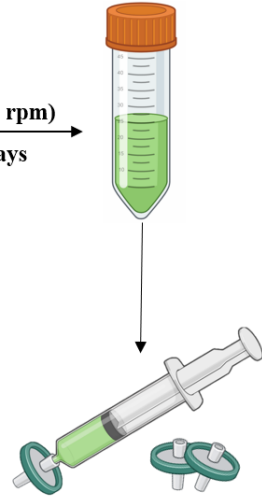
PDB medium

5 mm mycelial plugs of *Trichoderma* spp.



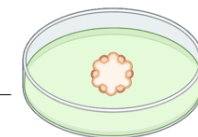
Rotary agitation (150 rpm) at 25 ± 1°C for 7 days

Centrifugation at 5000 rpm for 20 min



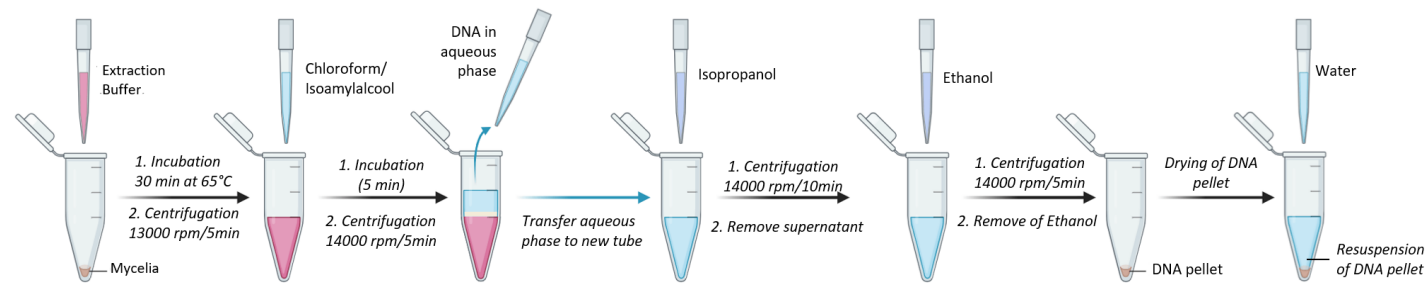
Introduced into sterilized PDA medium

Incubation for 6 days at 25°C



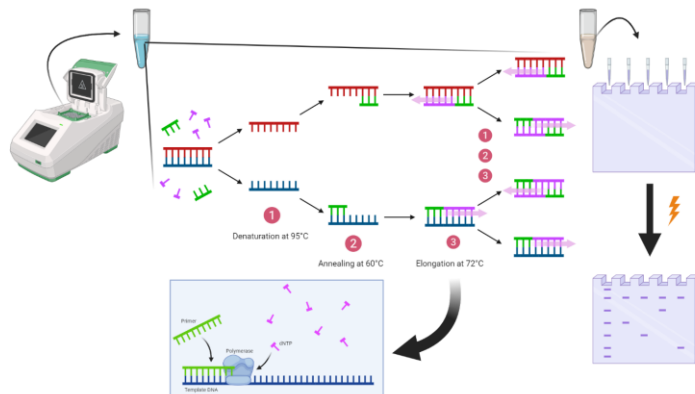
## Molecular identification

### 1 DNA EXTRACTION Doyle & Doyle (2007)

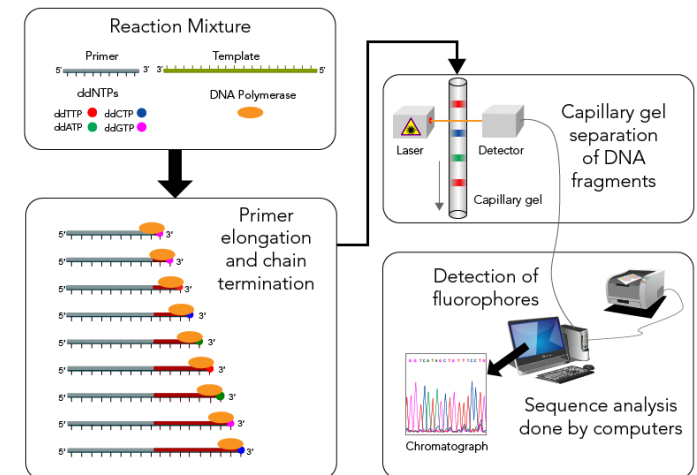


### 2 Polymerase Chain Reaction (PCR)

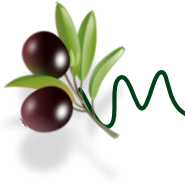
- Amplification of the internal transcribed spacer region ITS
- Primers (ITS1/ITS4)



### 3 Sequencing



# Methodology



## Biochemical characterization

### Qualitative screening of extracellular enzymes

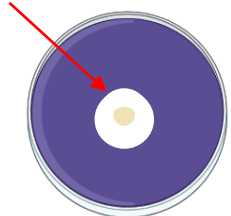
#### Amylase

Starch-based medium



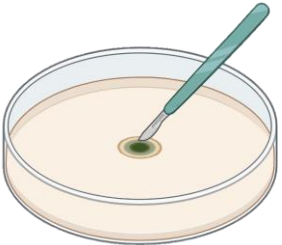
Lugol (0,2% KI)  
12, 2%

clear halos surrounding the colonies



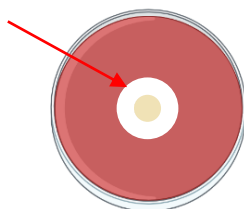
#### Cellulase

CMC-based medium



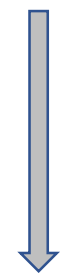
Congo red (0,1%)

clear halos surrounding the colonies

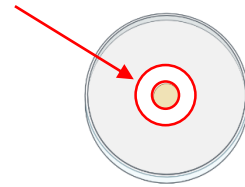


#### Protease

skim milk-based medium

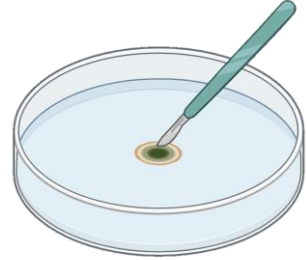


clear halos surrounding the colonies

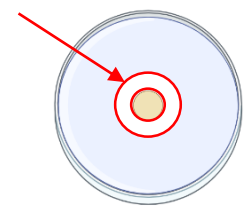


#### Chitinase

Chitine-based medium



clear halos surrounding the colonies





# Methodology



## Application

### In vivo assay

#### Treatments applied in the greenhouse bioassay

| Treatments | Composition            | Repetition | Period   |
|------------|------------------------|------------|----------|
| T-MK1      | SS + pathogen + T-MK1  | 8          | 3 months |
| T-BM6      | SS + pathogen + T-BM6  |            |          |
| T-BM13     | SS + pathogen + T-BM13 |            |          |
| T-KH4      | SS + pathogen + T-KH4  |            |          |
| T-BK9      | SS + pathogen + T-BK9  |            |          |
| T-CS6      | SS + pathogen + T-CS6  |            |          |
| T-E8       | SS + pathogen + T-E8   |            |          |
| T-E11      | SS + pathogen + T-E11  |            |          |
| C+         | SS + pathogen          |            |          |
| C-         | SS only                |            |          |

\*SS: sterile soil

#### 1 Soil sterilization

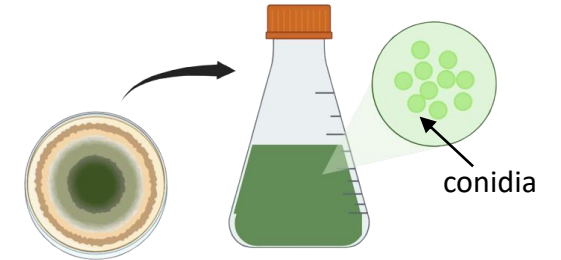


- 121 °C
- 90 min (2 times)



#### 2 Treatment preparation

200 mL *Trichoderma's* conidial suspension



#### 3 Inoculum preparation

50 g of wheat seeds



Pathogen + 50 mL of V8 jus

Incubation at 25°C/3 weeks

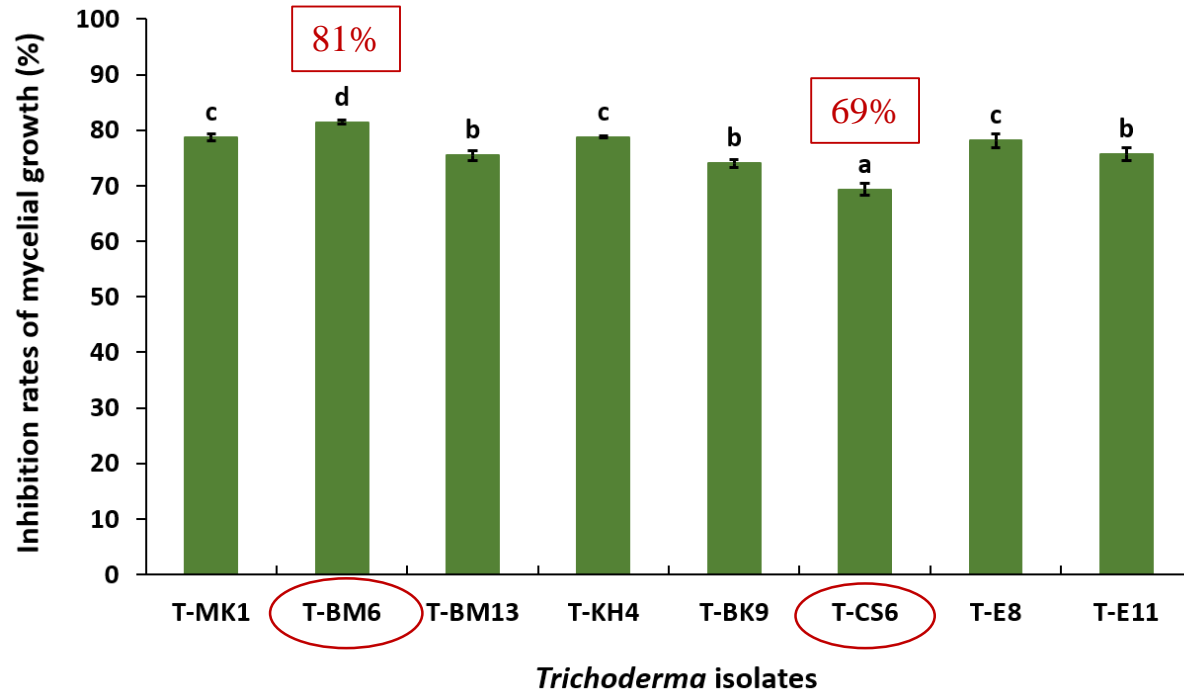
#### 4 Greenhouse assay



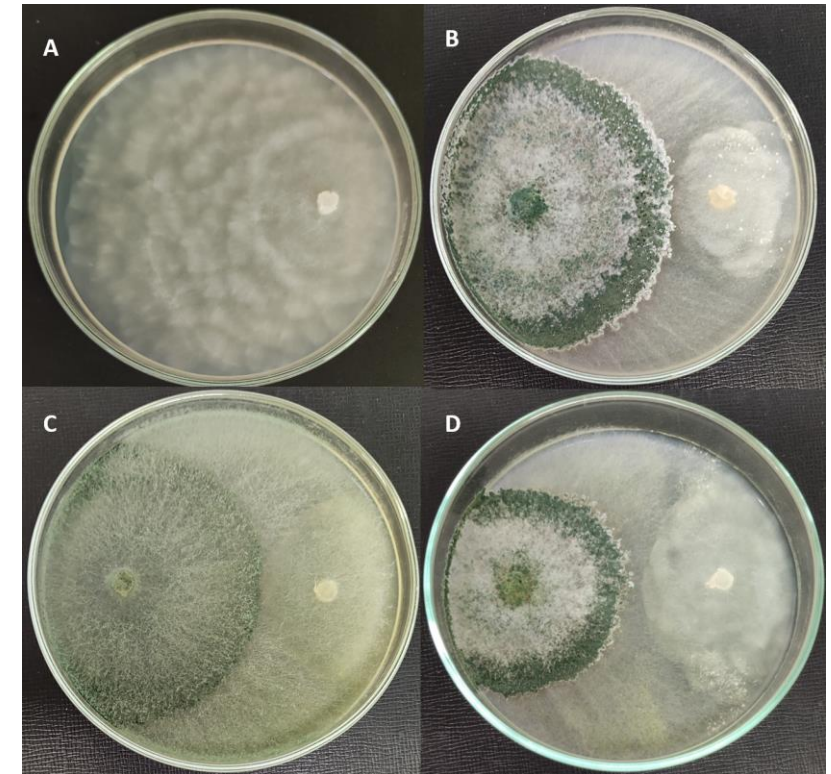
# Results



## Direct antagonistic activity



*In vitro* inhibition rates (%) of mycelial growth of *P. schmitteneri*, obtained with eight selected *Trichoderma* isolates from olive roots.



(A) control; (B) T-BM6; (C) T-BM13; (D) T-CS6

# Results



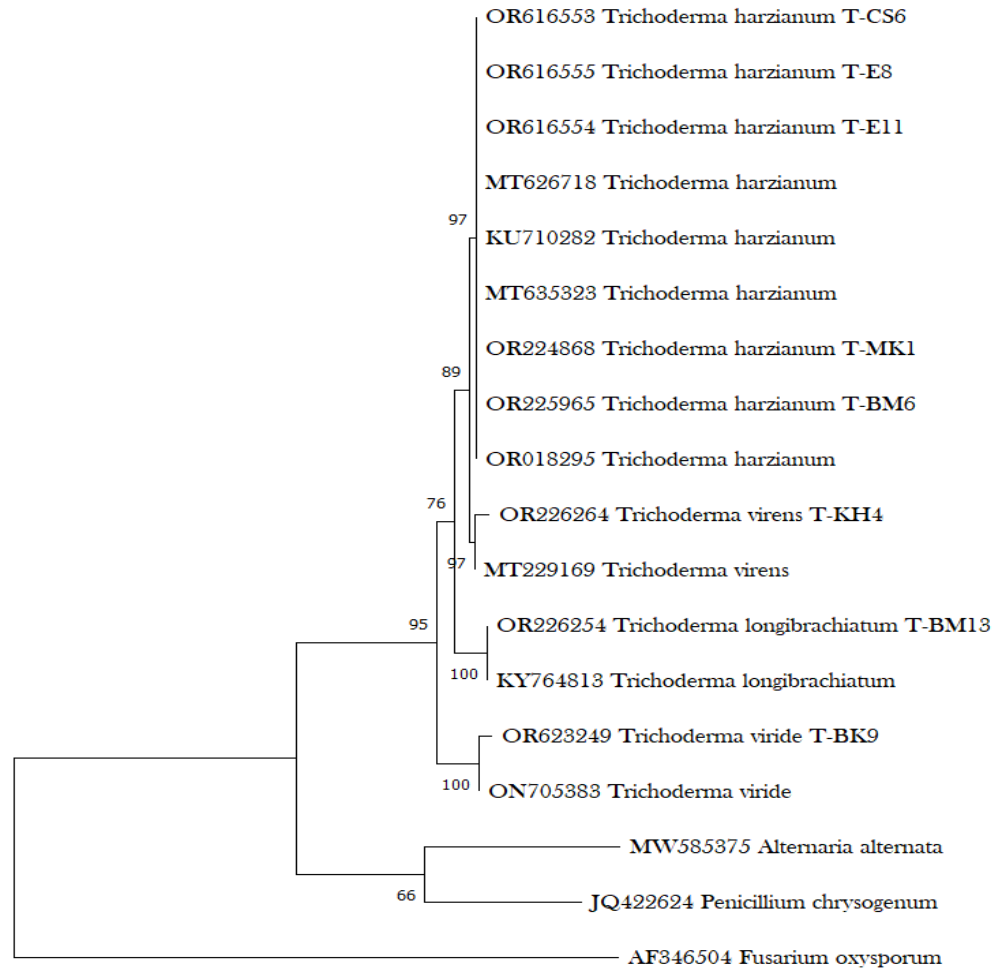
## Indirect confrontations

Effect of Volatile organic compounds (VOCs) and *Trichoderma* spp. filtrates (at 10% v/v) on the *in vitro* mycelial growth of *P. schmithenneri*

| Isolate code | Origin      | Species                  | Accession number | VOCs                      | Filtrate                  |
|--------------|-------------|--------------------------|------------------|---------------------------|---------------------------|
| T-MK1        | Meknes      | <i>T. harzianum</i>      | OR224868         | 65.65 ± 0.43 <sup>d</sup> | 54.77 ± 0.40 <sup>c</sup> |
| T-BM6        | Beni Mellal | <i>T. harzianum</i>      | OR225965         | 74.92 ± 0.45 <sup>f</sup> | 67.21 ± 1.57 <sup>e</sup> |
| T-BM13       | Beni Mellal | <i>T. longibrachatum</i> | OR226254         | 60.74 ± 0.37 <sup>c</sup> | 61.85 ± 0.45 <sup>d</sup> |
| T-KH4        | Khenifra    | <i>T. virens</i>         | OR226264         | 69.57 ± 0.68 <sup>e</sup> | 68.68 ± 1.26 <sup>e</sup> |
| T-BK9        | Beni Mellal | <i>T. viride</i>         | OR623249         | 57.46 ± 0.54 <sup>b</sup> | 44.41 ± 0.66 <sup>b</sup> |
| T-CS6        | Bouznika    | <i>T. harzianum</i>      | OR616553         | 49.71 ± 0.15 <sup>a</sup> | 44.61 ± 1.03 <sup>b</sup> |
| T-E8         | Errachidia  | <i>T. harzianum</i>      | OR616555         | 49.63 ± 1.24 <sup>a</sup> | 41.44 ± 0.65 <sup>a</sup> |
| T-E11        | Errachidia  | <i>T. harzianum</i>      | OR616554         | 56.67 ± 0.93 <sup>b</sup> | 40.52 ± 0.27 <sup>a</sup> |

# Results

## Molecular identification



- *Trichoderma harzianum* (5 isolates)
- *Trichoderma longibrachiatum* (1 isolate)
- *Trichoderma virens* (1 isolate)
- *Trichoderma viride* (1 isolate)

0.10

# Results

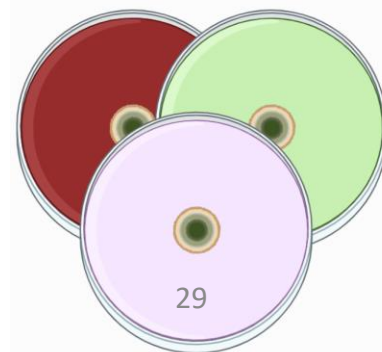


## Biochemical characterization

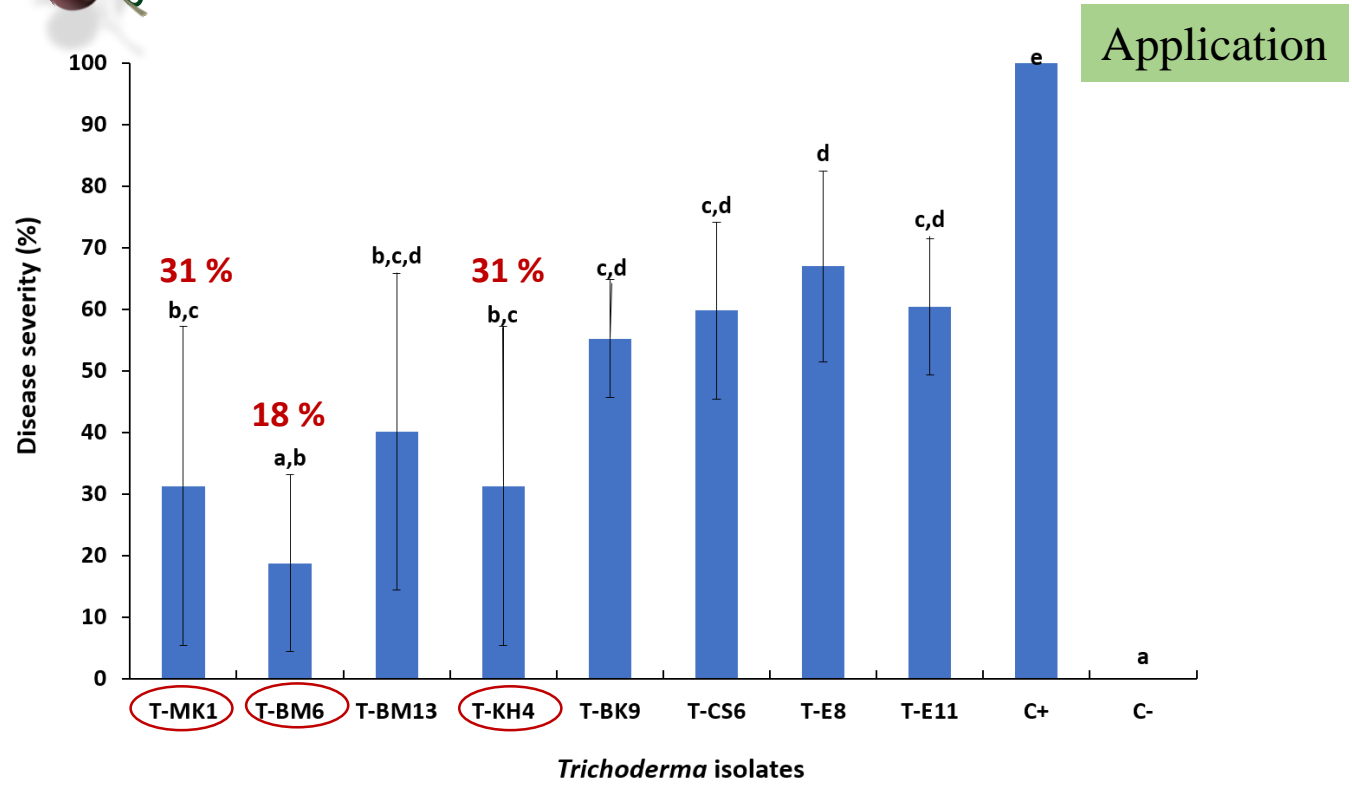
### Qualitative primary screening of *Trichoderma* spp. for enzymatic activity

| Isolate code | Amylase | Cellulase | Protease | Chitinase |
|--------------|---------|-----------|----------|-----------|
| T-MK1        | +++     | +         | -        | +         |
| T-BM6        | +++     | +++       | +        | +         |
| T-BM13       | +       | +         | -        | -         |
| T-KH4        | ++      | +         | +        | +++       |
| T-BK9        | +       | ++        | +        | +         |
| T-CS6        | ++      | +         | +        | +         |
| T-E8         | ++      | +         | +        | +         |
| T-E11        | ++      | +         | +        | +         |

(-): no enzyme activity; (+): very low enzyme activity; (++) : low enzyme activity; (+++) : high enzyme activity.



# Results



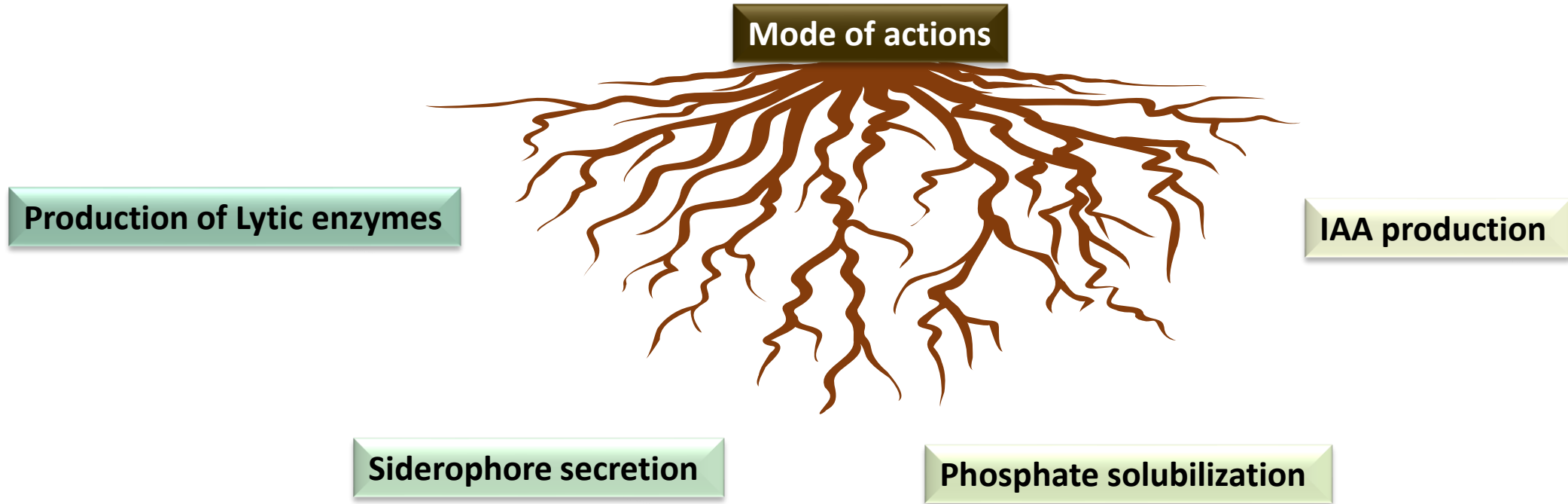
❖ *T. harzianum* T-BM6, *T. harzianum* T-MK1, & *T. virens* T-KH4 exhibited higher effectiveness in disease suppression in comparison to the positive control





# Conclusion and Perspectives

Screening and identification of 12 bacterial isolates with significant antifungal activity against the olive root rot pathogen

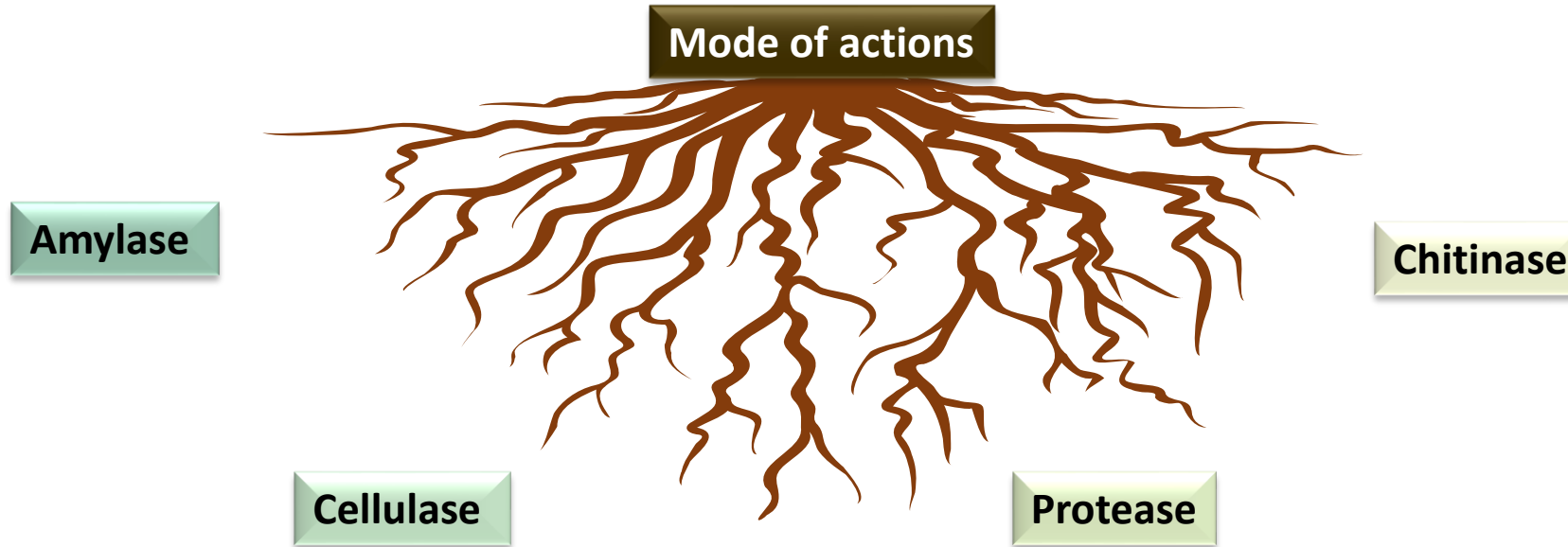


***Pseudomonas Koreensis A28* and *Bacillus subtilis C6* : Higher antifungal activity in vitro and in vivo**



# Conclusion and Perspectives

Screening and identification of 8 *Trichoderma* isolates with significant antifungal activity against *Pythium schmithennei*



*Trichoderma harzianum* T-BM6 : Higher antifungal activity in vitro and in vivo



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***Thank you for your  
attention***



***LEGRIFI Ikram***

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